**Abstract**

A new and simple method was developed to determine the stability constants of phenylalanine complexes of Co(II), Ni(II), Cu(II), Zn(II) and Pb(II) metal ions in water and water-dioxane mixtures at four different temperatures of 25, 37, 45 and 55°C potentiometrically using modified Bjerrum method. Ionic strength of medium was retained at 0.10 M by sodium nitrate. Our results revealed that the stability constant values are greater in water-dioxane mixtures than in water alone. The increasing trend in stability constant values in water and mixture of water-dioxane are the same as follows:

\[
K_{\text{Co(II)}-\text{Ph}} < K_{\text{Zn(II)}-\text{Ph}} < K_{\text{Cu(II)}-\text{Ph}} < K_{\text{Ni(II)}-\text{Ph}} < K_{\text{Pb(II)}-\text{Ph}}
\]

Furthermore, by knowing the stability constants at different temperatures, thermodynamic parameters of \(\Delta H^\circ\), \(\Delta S^\circ\) and \(\Delta G^\circ\) for the respective complexes were acquired. \(\Delta H^\circ\), \(\Delta S^\circ\) values were positive. Negative \(\Delta G^\circ\) values conveyed the spontaneity of the complex formation process. Also it is found out that the stability constant of the pertinent complexes increases as the temperature rises meaning that the reactions are endothermic.

**Keywords:** Stability constant, Amino acid complex, Potentiometric titrations, Thermodynamic parameters.

**Introduction**

The amino acids have special importance among the other chemical groups since they are building block of proteins. The interactions between metal ions and amino acids have attracted the attention of many biochemists, because they can be used as a model for metal-protein reactions mimicking metal-enzyme mechanism. The explanation of these phenomena in the biological systems requires the determination of the stability constants as a measure of how well the complex of the amino acids with various metal ions in a medium similar to those of biological systems forms.

Among various methods for determining stability constants of complexes, potentiometry has its own advantages. Potentiometric titration of amino acids in the presents of metal ions is generally used as a method for measuring metal complex stability constants. This technique first described by Bjerrum [1] and has been investigated extensively by numerous researches [2-7]. D.J. Perkins examined amino acid structures on the stabilities of complexes formed with metals of group II [8]. A. E. Martell and coworkers have conducted vital studies on amino acid complexes and predicted their stability constants [9-11].

The behavior of the complexes at different temperatures was probed by M.S. Masoud et al. [12]. Thermodynamic parameters for the formation of glycine with metal ions were investigated by S. Sammartano [13]. Formations of binary and ternary complexes were studied by M.M. Shoukry et al. [14]. Cu(II) amino acids complexes are useful antibacterial agents [15]. The stability constants of copper(II) complexes with several amino acids were calculated in dioxane-water mixtures by A. Dogan et al. [16].

The stability of binary complexes of L-aspartic acid in dioxane-water mixture was probed by R.S. Rani et al. [17]. H. Demirelli, et al. have determined the formation constants of phenylalanine complexes of Ni(II), Cu(II), and Zn(II) in water media at 25°C and \(\mu = 0.1 \text{ mol L}^{-1}\) KCl [18]. A.A. Mohamed et al. [19] have measured stability constants and thermodynamic parameters for glycine and L-threonine complexes with some rare metal ions in water. The interactions of L-glutamic and L-aspartic acid with some metal ions has been probed by S.A.A Sajadi [20]. Critical survey of formation constants of phenylalanine with metal ions has been reported by L.D. Pettit [21]. A. Eid Fazary, et al. have investigated the protonation equilibria of \(\alpha\)-amino acids in water and dioxane mixtures [22]. The stability constants of Ni(II) with some amino acids were probed by N. Turkel [23].

Phenylalanine is one of the few amino acids that can directly affect brain chemistry by crossing the blood-brain barrier. Phenylalanine is used to cure depression, attention deficit-hyperactivity disorder (ADHD), Parkinson’s disease, chronic pain, osteoarthritis, rheumatoid arthritis, alcohol withdrawal symptoms, and a skin disease called vitiligo [24].
In this work, the stability constants of phenylalanine complexes of some divalent metal ions in water and water-dioxane mixtures at four different temperatures have been evaluated. In addition thermodynamic parameters of pertinent complexes have been determined.

**Experimental Section**

a. **Materials and procedure**

Phenylalanine with purity of 99%, the nitrate salts of Co(II), Cu(II), Zn(II), Ni(II) and Pb(II) (all pro-analysis), nitric acid (HNO₃), sodium hydroxide (NaOH), hydrochloric acid (HCl), perchloric acid (HClO₄) and sodium nitrate (NaNO₃) all were purchased from Merck and used as received. Deionized water was employed in all of the experiments. The pH potentiometric titrations were performed using Schott pH meter, Thermostat MLW16, glass cell, digital burette, and magnetic stirrer.

A special glass vessel (reactor) for potentiometric titrations was made which had a double wall with entries for combined glass electrode, nitrogen, and base from burette. Temperature inside the reactor was kept constant through circulation of water with an accuracy of ±0.1°C. A 25.00 mL solution mixture prepared so that it was 5.000×10⁻³ M with respect to phenylalanine, 3.000×10⁻² M with respect to the respective metal ions and 1.690×10⁻² M with respect to HClO₄. A sufficient amount of 0.10 M NaNO₃ was added to adjust the ionic strength. The solution was thermostatted to desired temperatures of 25, 37, 45 and 55°C and then titrated with an accurately standardized NaOH solution while the titrand constantly was purged. The pH meter was calibrated after each addition of titrant in 0.050 ML increments. The two electrodes used for measuring pH were glass electrode and calomel electrode. The pH meter was calibrated using Merck standard buffer solutions purchased from Merck and used as received. Deionized water perchloric acid (HClO₄) and sodium nitrate (NaNO₃) all were purchased from Merck and used as received. Deionized water was employed in all of the experiments. The pH potentiometric titrations were performed using Schott pH meter, Thermostat MLW16, glass cell, digital burette, and magnetic stirrer.

b. **Calibration of the Glass Electrode**

Calibration of the combined glass electrode and calomel electrode was performed in both acidic and alkaline regions by titrating a solution of 0.01 mol L⁻¹ with pH of 4.0, 7.0 and 9.0.

The Bjerrum’s pH titration procedure assumes the presence of the reacting species H⁺ as amino acid, HL as the monoprotonated amino acid, and L⁻ as the anion of amino acid

**The Method for determination of stability constant**

The presence of the reacting species H⁺ as amino acid, HL as the monoprotonated amino acid, and L⁻ as the anion of amino acid

\[
\begin{align*}
\text{H}_2\text{L}^+ & \rightarrow \text{HL}^{(aq)} + \text{H}^+^{(aq)} \\
K_{a1} & = \frac{[\text{HL}][\text{H}^+]}{[\text{H}_2\text{L}^+]} \\
K_{a2} & = \frac{[\text{L}^-][\text{H}^+]}{[\text{HL}]} \\
M^{2+} + \text{HL} & \rightarrow \text{ML}^+ + \text{H}^+ \\
K_{f1} & = \frac{[\text{ML}^+][\text{H}^+]}{[\text{M}^{2+}][\text{HL}]} \\
M^{2+} + \text{L}^- & \rightarrow \text{ML}_2 \\
K_{f2} & = \frac{[\text{ML}_2]}{[\text{M}^{2+}][\text{L}^-]} \\
\end{align*}
\]

Here \( K_{a1} \) and \( K_{a2} \) is the first and the second stability constants of the complexes. We define \( \overline{n} \) as:

\[
\overline{n} = \frac{\# \text{ of bond ligands}}{\text{total metal ion concentration}} = \frac{L_{\text{bound}}}{C_M} = \frac{L_{\text{total}} - L_{\text{free}}}{C_M}
\]

The concentration of free ligand is the sum of concentration of contained ligand species at different form, i.e.

\[
L_{\text{free}} = [\text{H}_2\text{L}^+] + [\text{HL}^+] + [\text{L}^-]
\]

The bound ligand concentration \( L_{\text{bound}} \) could then be estimated as:

\[
L_{\text{bound}} = L_{\text{total}} - L_{\text{free}}
\]

After rearrangement and substitutions we have:

\[
\overline{n} = \frac{T_{\text{H}_2\text{L}^-} - [\text{H}_2\text{L}^-] - [\text{HL}] - [\text{L}^-]}{T_{\text{M}^{2+}}}
\]

Then:

\[
\overline{n} = \frac{[\text{ML}^+] + 2[\text{ML}_2]}{[\text{M}^{2+}] + [\text{ML}^+] + [\text{ML}_2]}
\]

According to mass balance relation we have:

\[
T_M = [\text{M}^{2+}] + [\text{ML}^+] + [\text{ML}_2]
\]

\[
T_{\text{HL}} = [\text{HL}] + [\text{L}^-] + [\text{ML}^+] + 2[\text{ML}_2]
\]
\[
\left[ \text{ClO}_4^- \right] = T_{\text{HClO}_4} + 2 T_M \\
\]
\[
[ML^+] + 2[ML_2] = [Na^+] - T_{\text{HClO}_4} + [H^+] \\
\]
\[
\overline{n} = \frac{[Na^+] - [HClO_4^+] + [H^+]}{T_M} \\
\]
\[
[HL] = \frac{K_a(T_{HL^+} - \overline{n}T_M)}{K_a + [H^+]} \\
\]

From plot of \( p_{\text{HL}} \) versus \( \overline{n} \), the stability constants could be calculated.

\[
K_{f1} = \frac{1}{[HL]^{1/2}} \\
K_{f2} = \frac{1}{[HL]^{1/2}} \\
\]

All our calculations in this work were executed by GRC\( \beta \)eta computer-program developed in our lab. The software asks for a) initial volume of solution containing the amino acid, metal ion, and perchloric acid, b) the concentration of perchloric acid, c) the concentration of sodium hydroxide, d) the concentration of amino acid, and e) \( pK_a \) and \( pK_a \) of the amino acid in the specified medium and at desired ionic strength which we found them in literature. After insertion of the pertinent values, the software plots calculated pH (corrected pH) of the titrant solution versus the concentration of added standardized NaOH plus drawing two curves, one for a \( \overline{n} = 0.5 \) and the other for \( \overline{n} = 1.5 \). The intersection of the potentiometric titration curve with these two curves produces two points (Figure 1) whose corresponding pHs will be used to evaluate the respective stability constants of the metallic ion-amino acid complexes. Additionally, the software is capable of plotting first and second derivative of d-pH versus d-concentration of NaOH to clarify the end points. For each potentiometric titration approximately 4-7 mL of standardized sodium hydroxide was used.

Thermodynamic calculations were conducted as follows:

The Gibb’s free energy change, \( \Delta G^0 \), can be calculated from the equation below:

\[
\Delta G^0 = -RT \ln K_f \\
\]

By taking the derivative with respect to \( 1/T \) from both sides of equation (23) we have:

\[
\frac{d \ln K_f}{d \frac{1}{T}} = -\frac{1}{R} \left( \frac{d \Delta G^0}{d \frac{1}{T}} \right) \\
\]

\[
\frac{d \ln K_f}{d \frac{1}{T}} = -\frac{1}{R} \left( \frac{\Delta H^0 + \Delta G^0}{T d \frac{1}{T}} \right) \\
\]

\[
\frac{d \ln K_f}{d \frac{1}{T}} = -\frac{1}{R} \left( \Delta H^0 - T \Delta S^0 + T \Delta S^0 \right) = \frac{-\Delta H^0}{R} \\
\]

So:

\[
\frac{d \log K_f}{d \frac{1}{T}} = -\frac{\Delta H^0}{2.303R} \\
\]

Regarding equation (27), the plot of log \( K_f \) versus \( 1/T \) produces straight line with slope equals:

\[
\text{slope} = -\frac{\Delta H^0}{2.303R} \\
\]

Using Equation (28) enables us to calculate Enthalpy change. For calculating \( \Delta S^0 \) we have:

\[
\Delta G^0 = \Delta H^0 - T \Delta S^0 \\
\]

Knowing Gibbs free energy and enthalpy changes we can evaluate \( \Delta S^0 \)

\[
\Delta S^0 = \frac{\Delta H^0 - \Delta G^0}{T} \\
\]

**Results and Discussion**

As an example, the output of the software as demonstrated in Figure 1 is a plot of pH versus concentration of added standardized NaOH for Cu(II)-Phe complex in (70-30) % water - dioxane mixture solution at 25°C.
complexes with respective metal ions. As it is cleared, with increasing the stability of the complex, titration curve for Cu (II) inclines more toward the right. Table 1 represents the stability constants values of the phenylalanine complexes of Co (II), Ni (II), Cu (II), Zn (II) and Pb (II) in temperatures of 25, 37, 45 and 55°C in aqueous solution. The stability constants of the complexes in 70-30% (v/v) water-dioxane mixture have been shown in Table 2. The results indicate that the order of increasing stability constants in both media are the same and as follows:

\[ K_{\text{Co(II)-Phe}} < K_{\text{Zn(II)-Phe}} < K_{\text{Pb(II)-Phe}} < K_{\text{Ni(II)-Phe}} < K_{\text{Cu(II)-Phe}} \]

This stability trend is in agreement with Irving-William series [26], which is based on ionic potential of metallic ions. The more charge density, the more electrostatic forces appear between ligand and metallic ion causing an increase in stability constant (except Cu (II)). Also the stability constant of complexes is related to their stabilization energies. Cu (II) with d^9 configuration has the highest stability energy (Jahan-Teller effect) and Co (II) possesses the lowest stability energy among the first transition series. Pb(II) is located in fourth period and its stability constant cannot be compared with the others.

Potentiometric titration curves for Cu (II)-Phe complexes

![Potentiometric titration curves for Cu (II)-Phe complexes](image)

**Figure 2:** Potentiometric titration curves for the respective complexes at 25°C in water solution; series 1: free of metal ions, series 2: Co (II) ions, series 3: Zn (II) ions, series 4: Ni (II) ions, series 5: Pb (II) ions and series 6: Cu (II) ions.

Table 1: The Log of the stability constants values for the respective metal ion-Phe complexes in aqueous solution at four different temperatures

<table>
<thead>
<tr>
<th>Complex</th>
<th>Stability constants</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Ni(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Cu(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Zn(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Pb(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Complex</th>
<th>Stability constants</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Ni(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Cu(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Zn(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
<tr>
<td>Pb(II)-Phe</td>
<td>Log k&lt;sub&gt;1&lt;/sub&gt;, Log k&lt;sub&gt;2&lt;/sub&gt;</td>
</tr>
</tbody>
</table>

for 50-50%, 70-30% (v/v) and water alone have been shown in Figure 3. The more increase in the stability constant of a complex, the more its titration curve is drawn to the right. This means higher stability constant causes more H<sup>+</sup> to be released at lower pH. By changing the solvent, the acidity and basicity of solute varies. The acidic and basic dissociation constant of any species will be measured with respect to the solvent. If a solvent with dissociation constant value of less than water is used, the acidic property of that species increases, therefore the shape of titration curve inclines toward the lower pH with respect to water as solvent.

With increasing the percent of dioxane, the stability constant increases. Because the dissociation constant of amine group of phenylalanine is lower in dioxane than water, so, the stability constant should decrease. This statement is in contrast with the above results. The discrepancy can be explained by solvating
ability of ML, molecular species, which have more solvating ability in an organic solvent than in water. This is due to lower dielectric constant of dioxane, 2.3 with respect to water, 80. Instead, the solvating ability of M^{2+}-molecular ion species is higher in aqueous solution than in organic solvent. It can be expected that the stability constant values are greater in aqueous-organic mixture than in aqueous alone.

The thermodynamic parameters values in Tables 3 and 4 indicate that change in enthalpy for water and water-dioxane mixtures are positive, showing the reactions endothermocity. In all complex reactions with metal ions, the Gibb’s free energy changes are negative referring to the reactions spontaneity. The trend has the same pattern for the formation of the complexes in water and in water-dioxane mixtures. It is worthy to note that on increase in the dioxane contents, the free energy becomes more negative, which is an evidence for increasing the stability of the respective complexes.

**Conclusions**

The stability constants of some divalent metal ion-Phe complexes in water, 70-30% and 50-50% (v/v) water-dioxane mixtures have been determined. The results indicate that the least stable complex is Co (II)-Phe and the most stable one is Cu (II)-Phe. As the percentage of dioxane in the solvent mixture increases, the stability of complexes rises too. This is due to a decrease in dielectric constant of water with respect to dioxane. In fact, co-solvent could affect the protonation-deprotonation equilibria in solution. This will happen by change in dielectric constant of the medium, which alters the relative contribution of electrostatic and non-electrostatic interactions. Furthermore, thermodynamics parameters of ΔH°, ΔS° and ΔG° were calculated. The data shows that the enthalpy change is positive for all the complexes indicating the reactions are endothermic. The negative ΔG° values for all complexes gives an evidence for spontaneity of the complex reactions.

**Acknowledgement**

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**References**

14. Shoukry MM, Shehata MR, Mohamed MMA. Binary and ternary

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**Table 3:** Thermodynamic parameters for the pertinent metal ion – Phe complexes in water at 25°C

<table>
<thead>
<tr>
<th>Complex</th>
<th>ΔH° (KJ/mol)</th>
<th>ΔS° (J/K)</th>
<th>-ΔG° (KJ/mol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Co(II)-Phe</td>
<td>6.16</td>
<td>99.2</td>
<td>23.4</td>
</tr>
<tr>
<td>Ni(II)-Phe</td>
<td>26.7</td>
<td>198.6</td>
<td>32.5</td>
</tr>
<tr>
<td>Cu(II)-Phe</td>
<td>49.2</td>
<td>306.4</td>
<td>42.1</td>
</tr>
<tr>
<td>Zn(II)-Phe</td>
<td>44.85</td>
<td>269.2</td>
<td>35.4</td>
</tr>
<tr>
<td>Pb(II)-Phe</td>
<td>6.91</td>
<td>100.3</td>
<td>23.0</td>
</tr>
</tbody>
</table>

**Table 4:** Thermodynamic parameters for the pertinent metal ion – Phe complexes in water-dioxane mixture at 25°C

<table>
<thead>
<tr>
<th>Complex</th>
<th>(70-30%) (v/v) water-dioxane</th>
<th>(50-50%) (v/v) water-dioxane</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ΔH° (KJ/mol)</td>
<td>ΔS° (J/K)</td>
</tr>
<tr>
<td>Co(II)-Phe</td>
<td>20.2</td>
<td>16.2</td>
</tr>
<tr>
<td>Ni(II)-Phe</td>
<td>19.1</td>
<td>177.1</td>
</tr>
<tr>
<td>Cu(II)-Phe</td>
<td>16.8</td>
<td>213.5</td>
</tr>
<tr>
<td>Zn(II)-Phe</td>
<td>55.4</td>
<td>307.0</td>
</tr>
<tr>
<td>Pb(II)-Phe</td>
<td>9.90</td>
<td>125.3</td>
</tr>
</tbody>
</table>

**Table 5:** Comparison of the stability constants values for the pertinent metal ion - Phe complexes in water at 25°C in or Lab and in the literature

<table>
<thead>
<tr>
<th>Cation</th>
<th>Co²⁺</th>
<th>Ni²⁺</th>
<th>Cu²⁺</th>
<th>Zn²⁺</th>
<th>Pb²⁺</th>
</tr>
</thead>
<tbody>
<tr>
<td>log β</td>
<td>log β</td>
<td>log β</td>
<td>log β</td>
<td>log β</td>
<td>log β</td>
</tr>
<tr>
<td>Acquired in our Lab</td>
<td>4.22, 7.83</td>
<td>5.80, 9.97</td>
<td>7.57, 13.78</td>
<td>4.61, 9.17</td>
<td>5.69, 9.25</td>
</tr>
<tr>
<td>The literature [21]</td>
<td>4.08, 8.08'</td>
<td>5.46, 9.99</td>
<td>7.51, 14.25</td>
<td>4.80, 9.11</td>
<td>4.03, 8.79</td>
</tr>
</tbody>
</table>

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**Citation:** Ghiamati E (2017) Thermodynamic Studies on the Interaction between Phenylalanine with Some Divalent Metal Ions in Water and Water-Dioxane Mixtures. SOJ Biochem 3(1):1-6. DOI: http://dx.doi.org/10.15226/2376-4589/3/1/00121


