

A Messenger Ribonucleic Acid: A Platform for Protein Synthesis

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Abstract

The messenger Ribonucleic Acid (mRNA) has come to play as potential regulators of gene expression that is translates in the ribosome by the RNA polymerase and yields amino acid and protein. This plays a tremendous role in the protein synthesis and is not just mere template in translation but also contains essential regulatory elements as protein supplement, for therapy vaccine production and generation of pluripotent stem cells. This review highlights the new approaches and insights in the role of mRNA as a template in protein synthesis, functional approach, clinical uses and in vivo usage which is still a challenge underway.

Keywords: MRNA; Translation; Protein synthesis; DNA; Gene regulation.

Introduction

The mRNA is a single stranded RNA transcribed from the DNA in its orderly arranged manner that has two extreme ends of cap and tail, 5' and 3' untranslated regions (UTR) and coding sequence (CDS) in middle which translates to proteins. The amino acid synthesis starts in the coding region with the initiation codon AUG and runs to the termination codon AAA. The 5' terminal cap structure makes room for the correct binding between mRNA and ribosome. The ribosome is an enzymatic protein that catalyzes the mRNA for the translation process to yield protein [1]. The translation machinery is located in the ribosome which begins at the 5' end of the mRNA but the 3' end is still join to DNA. But for the protein synthesis, translation mechanism of mRNA is programmed in away to integrate the amino acid for its production [2]. In the 3'non coding region as the location of microRNAs (miRNAs) that regulate the gene expression. This also contributes to mRNA degradation and translational repression due to its interaction with mRNAs at 3'UTR mRNAs activate translation or regulate transcription [3]. However, the primary function of it is in protein synthesis. This mechanism happens in a way that the translation into protein occurs by the action of transfer RNA and ribosome and two major ribosomal RNA (rRNA) molecules [4]. This involves the breakdown of large and small subunits of ribosome by the initiation factors (I_fs), whereby

the small subunit (SSU) bounds to mRNA and met-transfer RNA and forms the initiation complex. It is followed by the peptide change elongation (PCE) that integrates the met-tRNA at the P site coupled with the amino acyl -tRNA to elongation factors (EFs) and binds at a site. The 5' and 3' UTR provides stability for the mRNA. Within the ribosome the mRNA initiates proteins at coding region [5]. The publication of Avery, McLeod and McCarty led to the recognition of DNA and its role in genetics due to the transformational principle of pneumococcal bacteria [5, 6]. Later, the mRNA came to a firm resolution for the knowledge of gene function [7]. In 1957, Arthur Pardee during his sabbatical visit to the Institute, Pasteur recognized that genes produce a messenger molecule as the first time in Paris [5]. It was later on that the RNA fraction was called messenger RNA (mRNA) by Jacob and Monod [6]. But this was debated that Nirenberg was the first to isolate mRNA [10].

Functional Approach to MRNA

Recently, mRNAs have come into play as potential regulators of gene expression following the RNA modifications in coding sequences N⁶ - methyl adenosine, (M⁶A), 5 - methyl cytosine (M⁵C); pseudouridine Y) and N¹ methyadenosine ((M¹A) have been discovered within open reading frames of mRNAs [12].

The modulation of mRNAs within a cell has helped in the direct mechanism of protein synthesis regulation. But, a link between the amount of mRNA and its corresponding protein has not been observed [13, 14].

The disruptions in the initiation factor in the protein synthesis is not the only factor but the structural motif and regulatory activities in the 5' or 3' untranslated regions (UTRs) of MRNA come into that[15 -18].

Furthermore, mRNAs are not just mere templates for translation, but contain essential regulatory elements which might be included in regulation of translation. Certainly, the modification of mRNA has opened way for N' methyladenosine (M¹A) to be included in the messenger RNA bank and this led to the elevated translation rates [18, 19]. This is known to be of utmost interest in the genetic information at the RNA level

which is Adenosine – Inosine editing [21, 22]. The issue of mRNA modification which contributes a lot in the translation mechanism acts as markers to provide landing platforms for proteins which enhance other regulatory processes like mRNA degradation or localization [23-28].

Following the systematic approach to the mechanism of mRNA in processing and transport has made impact in the gene expression and when located in sub cellular level also influence polarity, migration and development [29, 30].

However, the protein elongation and co translational protein folding as shown by mRNA also its particular structure in the regulation of translation rate, but the actual supports to these potential mechanisms to the control of translation speed remain uncertain and debatable [31 -37].

Notwithstanding, the shift in translational rates in the mRNA structure – dependent can bring changes thereby making enough protein that can cause major phenotypic impacts such as human disease [38].

Shabalina and colleagues went further in their studies that changes in translation speed regulated by secondary structure of mRNA has led to variations in post-translated modifications of the nascent polypeptide which was not earlier regarded in RNA level regulation [39].

Also, remarkable evidence is the mRNA-translation speed-change (MTSC) occurs during multi drug resistance I (MDR₁ or ABCB1) gene that attacks protein folding and changes the conformation, the structure and function of the multidrug transporter [40, 41].

This awareness of mRNA nucleotide modification has led to the invention of RNA mass spectrometry and the next generation sequencing (NGS) techniques which has broadened our horizon to understand the concept that M6A Pseudouridine(Y) in the mRNA is a part in biological role such as splicing and regulation of translation to mRNA decay [42,43].

Whereas the ribosomal associated ncRNA (rncRNAs) binds with mRNA and reduces the protein synthesis in stress dependent manner [44, 45].

The recent study has also led to the discovery of micro RNAs (miRNAs) [46] small RNA molecules (18-25 nucleotides), which in their nature single stranded forms that acts as post transcriptional repressors through pairing with messenger RNA [47]. Elgar and colleagues in their approach to the field of genetic engineering discovered that synthesized mRNA (Syn-mRNA) has been of much value and safe than the DNA in the genomic integration for human clinical therapy [48].

Although Wolf and others have shown in 1990 that direct injection of 'naked' messenger RNA (mRNA) into the skeletal muscle of a mouse resulted in expression of the encoded protein development of mRNA vaccines, but this was considered unrealistic because of the expected mRNA instability during storage and after application *in vivo* [49].

More recently, Warren and his colleagues have demonstrated the success of mRNA- mediated reprogramming of human fibroblasts to induce pluripotent stem cells (iPSC) and in turn, the mRNA mediated differentiation to achieve desired mature cell type [50].

UTRs are sequences present at the 5' and 3' ends of a transcript that do not code for proteins but carry features that allow differential regulation of a gene [51].

In humans, the disarrangement in this region of UTRs results in deregulation of genes which leads to susceptibility to diseases [52].

MRNA and Evolution

The recent discoveries have shown that the outcome of new genes is fundamental to the evolution of lineage- specific traits, and several molecular mechanisms regarding the origin of new genes [53-57]. Ohno proposed that new functions could be derived through DNA based gene duplications, and has gone extensive study as a major source of new genetic material [58, 59].

Another form of new gene formation is mRNA based retroposition where reverse transcription of mRNA generates new intronless retrocopies that are fractioned into random genomic regions [57, 60, 61]. The means of finding out the fundamental method of gene expression of evolution has been on the detection of cis-acting genomic regulatory elements and trans- acting regulatory factors that decodes differences between species and populations [62 - 65]. The regulatory changes caused by gene expression variations have been a factor in phenotypic differences between closely related species [69 - 71].

Meyer and Zhou discovered that 5' UTR M6A modification makes way for cap independent translation initiation that functions for selective mRNA translation mechanism [72, 23]. It is of universal concept that N6 methyladenosine M6A RNA transcript modification among high animals as discovered in genetic engineering [73-76], and its reversible nature [73, 76, 77, 79]. Its effects on mRNA life time and translation efficiency have its role in the regulations of key regulators of biological pathways in human disease [27, 28, 80, 81 and 75].

Conclusion

The MRNA has much impact in the field of genetics as a potential gene regulator and its modulation effects in protein synthesis. The mRNA modification acts as a marker in accessing a landing platform for proteins. A certain shift in translational rates in mRNA structure-dependent brings changes thereby increasing production which leads to human disease. However, the heterogeneous nature of mRNA has made it to be involved in genomic integration for human clinical therapy.

References

1. Biology Dictionary
2. Chapeville F, Lipmann F, Von Ehrenstein G, Bernard Weisblum, William

- J. Ray, Seymour Benzer. On the role of soluble ribonucleic acid in coding for amino acid, *Proc Natl Acad.* 1962; 48(6): 1086-1092. doi: 10.1073/pnas.48.6.1086
3. Jacob O'Brien, Heyam H, Yara Z and Chun P. Overview of microRNA biogenesis mechanisms of actions and circulation. *Experimental Endocrinology*.2018; 9: 402. doi:10.3389/fendo.2018.00402
4. Griffiths A. J. F, Jeffrey H. Miller, David T. Suzuki, Richard C. Lewontin and William M. Gelbart Freeman. *Three Roles of RNA in Protein Synthesis. An Introduction to Genetic Analysis* 5th edition. 1993; 9(11):399. doi:10.1016/0168-9525(93)90141-4
5. Jacob, F and Monod J. Genetic regulatory mechanisms in the synthesis of protein. *Journal of Molecular Biology* .1961; 3(3): 318-356.doi: 10.1016/S0022-2836(61)80072-7
6. Avery O. T., Macleod, C.M and Mc Carty, M. Studies on the chemical nature of the substance inducing transformation of Pneumococcal types. Induction of transformation by a deoxyribose nucleic acid traction isolated from Pneumococcus types iii. *Journal of Experimental Medicine*.1944; 79(2):137-158.
7. Cobb M. Essay: - Who discovered messenger RNA? *Cell Press Current Biology*. 2016; 25(13): R523- R548.doi: 10.1016/j.cub.2015.05.032
8. Pardee AB. Paja Mas in Paris. *Trends Genet*.2002; 18(11): 585-587.
9. Jacob, F and Monod J. Genetic regulatory mechanisms in the synthesis of protein. *Journal of Molecular Biology*.1961; 3:318-356.
- 10.Starkey Lewis, P.J, Dear, J., Platt V., Costello, E.M Neoptolemos, D.J. French, N.S. Circulating micro RNAs as potential markets of human drug- induced liver injury. *Hepatology*. 2011; 54(5):1767-1776. doi: 10.1002/hep.24538
- 11.Xu, L., Liu,X, Sheng, N, Oo, K S., Liang, J Chionh, YH. Three distinct 3- methyl cytidine (M3C) methyl transferases modify tRNA and mRNA in mice and humans. *Journal of Biological Chemistry*.2017; 292(35):14695-14703. doi: 10.1074/jbc.M117.798298
- 12.Thomas, P.H., Alexander H, Mathias D.E. In RNA modifications Dynamic regulations of gene expression? *RNA Biology*.2016; Vol. 13 9: 760-765
- 13.Maier T, Güell M, Serano L. Correlation of mRNA and protein in complex biological samples. *FEBS let* .2009; 583(24): 3966-73. doi: 10.1016/j.febslet.2009.10.036
- 14.Taniguchi Y, Choi Pj, Li GW, Chan H, Babu M., Hearn J. Quantifying E. Col, Proteome and transcriptome with single-molecule sensitively in single cells. *Science*.2010; 329(5991): 533-8. doi: 10.1126/science.1188308
- 15.Gebauer F, Hentze MW. Molecular mechanisms a translation control. *Nature Review Molecular Cell Biology* 2004; 5(10): 827-35. doi:10.1038/nrm1488
16. Xue S, Barna M. Specialized ribosomes: a new frontier in gene regulation and organismal biology. *Nature Review of Molecular cell Biology* .2012; 13(6): 355-369. doi: 10.1038/nrm3359
- 17.Xue S, Barna M . Cis- regulatory RNA elements that regulate specialized ribosome activity. *RNA Biology*. 2015; 12(10): 1083-1087. doi: 10.1080/15476286.2015.1085149
- 18.Sonenberg N, Hinnebusch AG. Regulation of translation initiation in eukaryotes: mechanisms and biological targets. *Cell*. 2009; 136(4): 731-45. doi:10.1016/j.cell.2009.01.042
- 19.Dominissini D, Nachtergele S, Moshitch Moshikiritz S, Peer E, Kol N, Ben- Haem M. S, Dai Q, Di. The dynamic N (1) - methyladenosine, methylome in eukaryotic messenger RNA. *Nature*.2016; 12: 311-316.
- 20.Li X, Xiong X, Wang K, Wang L, Shu X, Ma S. Transcriptome wide mapping reveals reversible and dynamic N -1 methyl adenosine methylome. *Nature Chemical Biology*.2016; 12:311-316. doi: 10.1038/nchembio.2040
- 21.Nishikura K, A-to I editing of coding and non- coding RNAs by ADARS. *Nature Reviews in Molecular cell Biology*.2016; 17: 83-96.
- 22.Mallela Nishikura K. A-to I editing of protein coding and non coding RNAs. *Critical Reviews in Biochemistry and Molecular Biology*.2012; 47(60): 493-501. doi:10.3109/10409238.2012.714350
- 23.Zhou J, Wan J, GaoX, Zhang X, Jaffrey SR, Qian SB .Dynamic m(6)A mRNA methylation directs translational control of heat shock response. *Nature* .2015; 526(7574): 591-594. doi: 10.1038/nature15377
- 24.Wang X .Zhao BS, Roundtree IA, Lu Z, Han D, Ma H. N(6)- methyl adenosine modulates messenger RNA translation efficiency. *Cell* .2015; 161(6): 1388- 1399.doi: 10.1016/j.cell.2015.05.014
- 25.Xu C, Wang X. Liuk, Roundtree I.A, Tempel W, Li Y . Structural basis for selective binding of m6A RNA by the YTHDC1 YTH domain. *Nature Chemical Biology*. 2014; 10(11): 927-929. doi: 10.1038/nchembio.1654
- 26.Liu N, Dai Q, Zheng, He C, Parisien M, Pan T. N (6) - methyladenosine dependent RNA structure switches regulates RNA protein interactions. *Nature* .2015; 518(7540): 560-564. doi: 10.1038/nature14234
- 27.Wang X, Lu Z, Gomez A, Hon GC, Yue Y, Han D. N6- methyl adenosine-dependent regulation of messenger RNA stability. *Nature*. 2014; 505(7481):117-120.doi: 10.1038/nature12730
- 28.Fustin JM., Dio, M., Yamaguchi Y, Hida H, Nishimura S, Yoshida M. RNA methylation- dependent RNA processing controls the speed of the circadian clock. *Cell*. 2013; 155(4): 793-806. doi:10.1016/j.cell.2013.10.026
- 29.Parton, R.M, Davidson, A., Davis I., and Weil, T.T. Subcellular mRNA localization at a glance *Journal of Cell. Biology*. 2014; 127:2127-2133. doi: 10.1242/jcs.114272
- 30.Buxbaum.A.R, Haimovich, G., and Singer R.H. In the right place at the right time: visualizing and understanding mRNA localization. *Nature Reviews in Molecular Biology Cell*.2015; 16(20):95-109. doi:10.1038/nrm3918
- 31.Gingold, H, and Pilpel Y. Determinants of translation efficiency and accuracy. *Molecular Systems Biology*.2011; 7: 481. doi: 10.1038/msb.2011
- 32.Shalgi R, Hurt J. A, Krykbaeva, I. Taipale, M, Lindquist, S , Burge C. B. Wide spread regulation of translation by elongation pausing in heat shock. *Molecular cell* .2013; 49(3):439-452. doi: 10.1016/j.molcel.2012.11.028
- 33.Richter, J.D. and Collier J. Pausing on polyribosomes: Make way for Elongation in translational control. *Cell* .2015; 163(2): 292-300. doi: 10.1016/j.cell.2015.09.041
- 34.Guilhem F. Aleksey Y, Svetlane A.S and Eugene V.K. Role of mRNA structure in control of protein folding. *Nucleic Acid Research* .2016; 44(22):10898-10911. doi: 10.1093/nar/gkw671
- 35.Artieri, C G. and Fraser H. B. Accounting for biases in ribo profiling data indicates a major role for proline in stalling translation. *Genome Reviews*.2014; 24(12): 2011- 2021. doi: 10.1101/gr.175893.114
- 36.Koutmou, K. Schuller, A.P. Brunelle J. I. Radhakrishnan A. Djuranovic, S and Green R. Ribosomes slides on lysine- encoding homopolymeric A stretches. *Elife* .2015; 4.doi: 10. 7554/ e life 05534

37. Requião, R. D, de Souza, H. J, Rossetto, S., Domitrovic T. and Palhano F. L. Increased ribosome density associated to positively changed residues is evident in ribosome profiling experiments performed in the absence of translation inhibitors RNA Biology.2016; 13(6): 561-568. doi:10.1080/15476286.2016.1172755
38. Chaney J L. And Clark P.L. Roles for Synonymous Codon Usage in Protein Biogenesis. Annual Review of Biophysics. .2015; 44: 143-166. doi: 10.1146/annurev-biophys-060414-034333
39. Shabalina, S. A, Spiriclonov N. A and Kashina, A. Sounds of Silence: Synonymous nucleotides as a key to biological regulation and complexity. Nucleic Acids Reviews .2013; 41(9): 2073- 2094. doi:10.1093/nar/gks1205
40. Kimchi-Sarfaty, C, Oh J. M, Kim, M. I W, Sauna, Z.E, Caleagno, A.M. A 'Silent' Polymorphism in the MDRI gene changes substrate specificity. Science. 2007; 315(5811):525-528. doi: 10.1126/science.1135308
41. Fung, K. L. Pan, J, Ohnuma .S, Lund, P.E, Pixley J. N. Kimchi- Sarfaty C. MDRI synonymous Polymorphisms alter transporter specificity and protein stability in a stable epithelial monolayer . Cancer Research. 2014; 74(2): 598-608. doi: 10.1158/0008-5472.CAN-13-2064
42. Wang X, Zhao B S, Roundtree I A, Lu Z, Han D, Ma H. N (6) - methyl adenosine modulates messenger RNA translation efficiency. Cell. 2015; 161(6): 1388-1399. doi: 10.1016/j.cell.2015.05.014
43. Zhao X, Yang Y, Sun BF, Shi Y, Yang X, Xiao W. FTO dependent demethylation of N6- methyl adenosine regulates mRNA splicing and is required for adipogenesis. Cell Reviews. 2014; 24(12):1403-1419. doi: 10.1038/cr.2014.151
44. Pircher A, Bakowska, Zywicka K, Schneider I, Zywicki M, Polacek N. An mRNA derived non coding RNA targets and regulates the ribosome. Mol Cell.2014; 54(1):147-155. doi: 10.1016/j.molcel.2014.02.024
45. Gebetsberger J, Zywick M, Kunzi A, Polack N. t RNA derived fragments target the ribosome and function as regulatory non coding RNA in *Haloferax volcanii*. Archaea. 2012; 11. doi: 10.1155/2012/260909
46. Lo, Y.M. Circulating nucleic acids in plasma and serum: an over view. Annals of the New York Academy of Sciences.2001; 945: 1-7. doi: 10.1111/j.1749-6632.2001.tb03858.x
47. Bartel, D.P. Micro RNAs: Target recognition and regulatory functions. Cell.2009; 136(2): 215-233. doi: 10.1016/j.cell.2009.01.002
48. Elgar, S and Guido K. Synthetic MRNAs for manipulating cellular phenotypes. An over view New Biotechnology 2015; 32(1): 1871-6784. doi: 10.1016/j.nbt.2014.04.008
49. Wolff JA, Malone RW, Williams P. Chong W, Aesadi G, Jani A. Direct gene transfer into mouse muscle in vivo. Science .1990; 247 (4949 Pt 1): 1465-1468. doi: 10.1126/science.1690918
50. Warren I, Manos P D, Ahfeldt T, Loh Y. H, Lau F. Highly efficient reprogramming to pluripotency and directed differentiation of human cells with synthetic modified mRNA. Cell Stem Cell.2010; 7(5):618-630. doi: 10.1016/j.stem.2010.08.012
51. Shelpa J R, Sangeeta C, Jayanta K. P. Untranslated regions of mRNA and their role in regulation of gene expression in protozoa parasites. Journal of Biosciences .2017; 42(1): 189-207.
52. Chatterjee S., and Pal, J K. Role of 5' and 3' untranslated regions of mRNAs in human disease. Biology of the cell.2009; 101(5): 251-262. doi: 10.1042/BC20080104
53. Burki, F and Kaessmann. Birth and adaptive evolution of a hominoid gene that supports high neurotransmitter flux. Nature Genetics.2004; 36 (10):1061-1063. doi: 10.1038/ng1431
54. Bradley J, Baltus A, Skaletsky H, Royce-Tolland M, Dewar and Page D C. A X to autosome retrogene is required for spermatogenesis in mice Nature Genetics.2004; 36(8): 872-876. doi: 10.1038/ng1390
55. Zhang G, Dahl JA , Niu Y, Fedoresak P, Huang CM, Li C J. ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. Molecular Cell.2013; 49(1):18-29. doi:10.1016/j.molcel.2012.10.015
56. Paulding CA. Ruvolo M. Haber D a. Tre-2(USP6) oncogene is a hominoid-specific gene. Proceedings of the National Academy of Sciences USA 2003; 100(5):2507-2511. doi: 10.1073/pnas.0437015100
57. Long M, Betran E, Thornton K, Wang W. The origin of new genes: glimpse from the young and old .Nature Reviews in Genetics.2003; 4:865-875.
58. Ohno S. Evolution by Gene Duplication. Springer-Verlag. Berlin 1970
59. Zhou Q, Zhang G, Zhang Y, Xu S, Zhao R, Zhan Z . On the origin of new genes in *Drosophila*. Genome Research.2008; 18(9):1446-1446. doi: 10.1101/gr.076588.108
60. Brosius J. Retroposons-seeds of evolution. Science. 1991; 251(4995):753. Doi: 10.1126/science.1990437
61. Kaessmann H, Vinckenbosch N, and Long M. RNA -based gene duplication, mechanistic and evolutionary insights. Nature Reviews in Genetics.2009; 10 (1):19-31. doi: 10.1038/nrg2487
62. Schmidt D, Wilson MD, Ballester B, Schwalie PC, Brown GD, Marshall A. Five vertebrate Chip- Seq reveals the evolutionary dynamics of transcription factor binding. Science.2010; 328(5981):1036-1040. doi:10.1126/science.1186176
63. Zheng W, Zhao H, Mancera E, Steinmetz LM, Snyder M. Genetic analysis of variation in transcription factor binding in yeast. Nature.2010; 464(7292): 1187-1191. doi: 10.1038/nature08934
64. He Q, Bradet AF, Patton B, Purvis J, Johnson J, Gogol M. High conservation of transcription factor binding and evidence of combinatorial regulation across six *Drosophila* species. Nature Genetics. 2011; 44:414-420.
65. Ni X, Zhang YE, Negre N, Chen S, Long M, White KP. Adaptive evolution and the birth of CTCF binding sites in the *Drosophila* genome. PLOS Biol.2012; 10: doi: 10.1371/journal.pbio.1001420
66. Mc Vicker G, Van de Geijn B, Degner JF, Cain CE, Banovich NE, Raj A. Identification of genetic variants that affect histone modifications in human cells. Science 2013; 342(6159):747-749. doi:10.1126/science.1242429
67. Arnold CD, Gerlach D, Spies D, Matts JA, Sytnikova YA, Pagani M. Quantitative genome- wide enhancer activity maps for five *Drosophila* species show functional enhancer conservation and turnover during Cis-regulatory evolution. Nature Genetics.2014; 46: 685-692.
68. Villar D, Berthelot C, Aldridge S, Rayner TF, Lukk M, Pignatelli M. Enhancer evolution across 20 mammalian species. Cell. 2015; 160(3): 554-566. doi: 10.1016/j.cell.2015.01.006
69. King MC, Wilson AC. Evolution at two levels in human and chimpanzees, Science.1975; 188(4184):197-116. doi: 10.1126/science.1090005

70. Romero IG, Ruvinsky I, Gilad Y. Comparative studies of gene expression and the evolution gene regulation. *Nature Reviews in Genetics*. 2012; 13:505-516.
71. Villar D, Flicek P, Odom DT. Evolution of transcription factor binding in metazoans-mechanisms and functional implications. *Nature Reviews in Genetics*. 2014; 15(4): 221-233. doi: 10.1038/nrg3481
72. Meyer KD, Patil DP, Zhou J, Zinoview A, Skabkin MA, Elemento O. 5-UTR M6A promotes cap-independent Translation. *Cell*. 2015; 163(4): 999-1010. doi: 10.1016/j.cell.2015.10.012
73. Dominissini D, Moshitch-Moshkovitz S, Schwartz S, Salmon-Divon M, Ungar L, Osenberg S. Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq. *Nature*. 2012; 485: 201-206.
74. Meyer KD, Saletore Y, Zumbo P, Elemento O, Mason CF, Jaffrey SR. Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. *Cell*. 2012; 149(7): 1635-1646. doi: 10.1016/j.cell.2012.05.003
75. Batista PJ, Molinie B, Wang J, Qu K, Zhang J, Li L. M6A RNA modification controls cell fate transition in mammalian embryonic stem cells. *Cell Stem*. 2014; 15(6): 707-719. doi: 10.1016/j.stem.2014.09.019
76. Luo GZ, MacQueen A, Zheng G, Duan H, Dore LC, Lu Z. Unique features of the m6A methylome in *Arabidopsis thaliana*. *Nature Communications*. 2014; 5:5630. doi: 10.1038/ncomms6630
77. Jia G, Fu Y, Zhao X, Dai Q, Zheng G, Yang Y. N6 methyladenosine in nuclear RNA is a major substrate of the obesity associated FTO. *Nature Chemical Biology*. 2011; 7:885-887.
78. Zheng G, Dahl JA, Niu Y, Fedorasak P, Huang CM, Li CJ. ALKB5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. *Molecular Cell*. 2013; 49(1): 18-29. doi: 10.1016/j.molcel.2012.10.015
79. Liu J, Yue Y, Han D, Wang X, Fu Y, Zhang L. A METTL3-METTL14 complex mediates mammalian nuclear RNA N(6)-adenosine methylation. *Nature Chemical Biology*. 2014; 10(2): 93-95. doi: 10.1038/nchembio.1432
80. Fustin JM, Do M, Yamaguchi Y, Hida H, Nishimura S, Yoshida M. RNA methylation-dependent RNA processing controls the speed of the circadian clock. *Cell*. 2013; 155(4): 793-806. doi: 10.1016/j.cell.2013.10.026
81. Schwartz S, Agarwala SD, Mumbach MR, Jovanovic M, Mertins P, Shishkin A. High-resolution mapping reveals a conserved, widespread dynamic in yeast meiosis. *Cell*. 2013; 155(6): 1409-1421. doi: 10.1016/j.cell.2013.10.047