

# Identification of Four Different Chlorophyll Allomers of *Nostoc* Sp. by Liquid Chromatography-Mass Spectrometer (LC-MS)

Bahareh Nowruzi<sup>1\*</sup> and Jouni Jokela<sup>2</sup>

<sup>1</sup>Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran

<sup>2</sup>Division of Microbiology, Department of Food and Environmental Sciences, University of Helsinki, Finland

Received: April 23, 2019; Accepted: May 14, 2019; Published: May 16, 2019

\*Corresponding author: Bahareh Nowruzi, Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran; E-mail: bahareh.nowruzi@srbiau.ac.ir

## Abstract

Cyanobacteria (Blue Green Algae) differ from other types of bacteria in that they have chlorophyll a, which other photosynthetic bacteria don't have. In this study, liquid chromatography-mass spectrometer (LC-MS) has been used for identification of the four different allomers of chlorophyll (Chlorophyll  $\alpha$ , HO-chlorophyll  $\alpha$ , HO-lactone-chlorophyll  $\alpha$  and MeO-lactone-chlorophyll  $\alpha$ ) from *Nostoc* sp. The differences in mass spectrometric fragmentation of Extracted ion chromatogram can be used as a diagnostic tool for the assignment of the configuration of four different chlorophyll allomers. This case is the first documented of identification of four different chlorophyll  $\alpha$  allomers from *Nostoc* sp. in Iran.

**Keywords:** chlorophyll  $\alpha$ ; allomers; *Nostoc*; liquid chromatography-mass spectrometer;

## Introduction

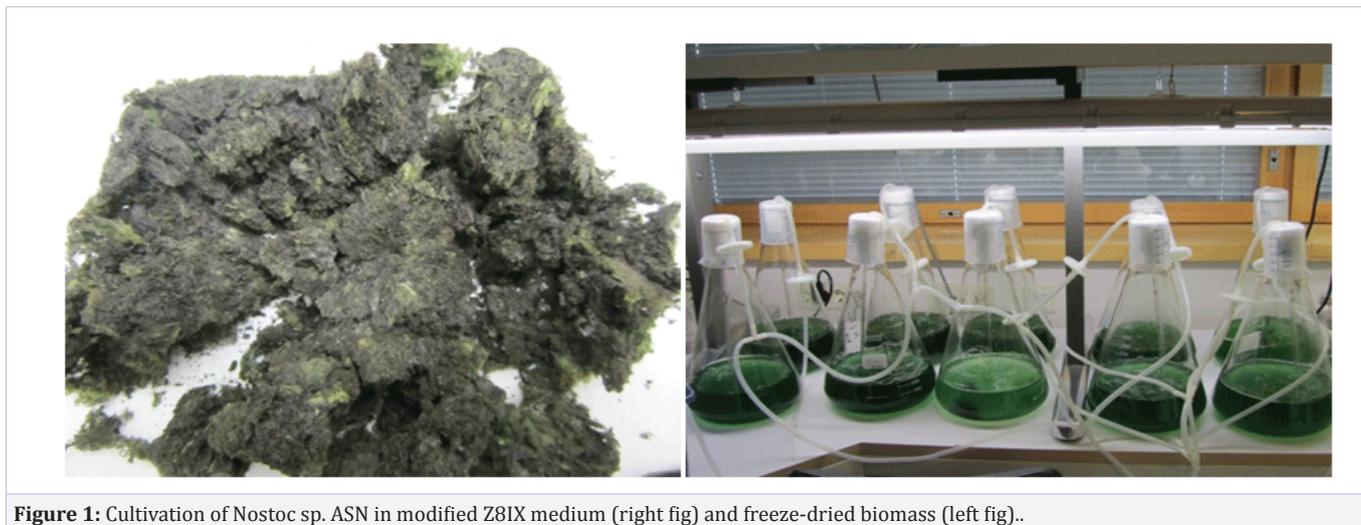
All plants, algae, and cyanobacteria which photosynthesize contain chlorophyll "a". Cyanobacteria contain only one form of chlorophyll, chlorophyll a, a green pigment. In addition, they contain various yellowish carotenoids, the blue pigment phycobilin, and, in some species, the red pigment phycoerythrin. The combination of phycobilin and chlorophyll produces the characteristic blue-green color from which these organisms derive their popular name. Because of the other pigments, however, many species are actually green, brown, yellow, black, or red [1]. The oxidation of the chlorophyll molecule by molecular triplet oxygen in alcoholic solutions causes the replacement of the atom of hydrogen of C-13 located in the isocyclic ring by oxygen or an oxygen-containing species [2]. This reaction named allomerization may occur by both enzymatic and chemical pathways, forming MeO-Lacton chl a, OH-chl a and MeO-chl a as major products [3]. The acid hydrolysis of the phytol alcohol in the chlorophyll molecule is accompanied by the loss of Mg and products pheophorbides. The excision of phytol without separation from Mg is a specific reaction catalysed by the endogenous enzyme chlorophyllase and which results in chlorophyllides [4, 5]. In Iran, algological studies are still scarce and limited to phylogenetic of genes encoding proteins

involved in bioactive compounds biosynthesis in paddy fields and fresh water regions [6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17 and 18]. Therefore, The objectives of this study was to develop a liquid chromatography-mass spectrometer (LC-MS) method for determination of Chlorophyll a and its derivatives in *Nostoc* sp. moreover we extend the use of MS/MS<sup>2</sup> in the designation of allomers of chlorophyll a configuration to identification of four different allomers during the methanolic allomerisation reactions of chlorophyll  $\alpha$ .

## Experimental Methods

### Strain cultivation and preparation of extract

*Nostoc* sp. ASN studied in this research was collected from paddy fields in Golestan province of Iran in 2010. It was grown at a photon irradiance of 15  $\mu\text{mol m}^{-2} \text{s}^{-1}$  in modified Z8IX medium for 21 days (Figure 1). The cells were harvested by centrifuge for 10 min at 10,000 g. After being lyophilized, the biomass was used for identification of the four different chlorophyll allomers of chlorophyll a. The extract for the chlorophyll a analysis was prepared from 1 ml of frozen culture. The microtube containing the culture was placed in a water bath (room temperature, 23°C), allowing the culture to thaw. Subsequently, it was supplemented with 300 mg of glass beads (disruptor beads, 0.5 mm, Scientific Industries) and 2  $\mu\text{l}$  of 50% (v/v) formic acid (Fluka, Sigma-Aldrich) (final concentration of 0.1% (v/v) formic acid). Proper cell disruption was achieved by placing the microtube into a homogeniser (FastPrep®-24 instruments, MP Biomedicals) for 15 s at a speed of 6.5  $\text{ms}^{-1}$ . The homogenised mixture was centrifuged at 10 000 G for 5 min. Twenty micro litres of the supernatant were diluted with 9 volumes of acetonitrile (E CHROMASOLV®, Sigma-Aldrich), equalling to a final volume of 200  $\mu\text{l}$ . The extract was analysed subsequently. For the preparation of methanol extracts, 236 mg of wet biomass were freeze-dried (Edwards's lyophilisator). The dry cell mass was measured to be 7.3 mg. A microtube containing the latter amount of dried cells, 300 mg of glass beads (disruptor beads, 0.5 mm, Scientific Industries) and 1 ml of methanol (LC-MS grade, Fischer Scientific) was placed into a homogeniser (FastPrep®-24 instrument, MP Biomedicals) for 15 s at a speed of 6.5  $\text{ms}^{-1}$ . The mixture was centrifuged as described above. The supernatant was stored refrigerated at 4°C



until chemical analyses were performed [19].

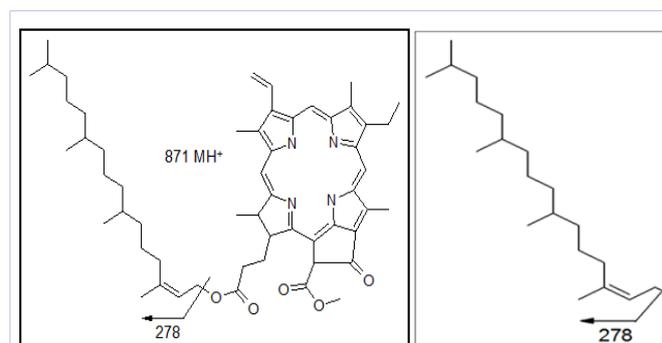
### Chemical Analysis

Nostoc sp. ASN cell extracts were analysed by liquid chromatography mass spectrometry (LC-MS) using an Agilent 1100 Series LC/MSD Trap XCT Plus System (Agilent Technologies). The sample was injected into the Luna C18 column (150X10 mm, 5 mm, Phenomenex) in batches (1 ml). The mobile phase A consisted of formic acid (0.1 percent) (Fluka, Sigma Aldrich, Steinheim, Germany) and mobile phase B consisted of Isopropyl alcohol. The injection volume of each sample was 10 µl. Setting of parameters for LC-MS has been shown as a (Table 1).

### Results and Discussions

The results of analysis by LC-MS showed the allomerization of chlorophyll a by forming MeO-Lacton chl a, OH-chl a and MeO-chl a as major products. The acid hydrolysis of the phytol alcohol in

the chlorophyll molecule is accompanied by the loss of Mg and produces chlorophyllides (Figure 2) (Table 2). Chromatogram in (Figure 3) showed mass spectrum of Total ion chromatogram (TLC) and Extracted ion chromatogram (EIC) of chlorophyll a molecule after separating of magnesium ion. The results of MS/MS<sup>2</sup> fragmentation pattern of two times protonated chlorophyll a allomers showed that in addition to magnesium, the phytol structure of chlorophyll molecule a has been isolated (Figure 4). Mass-to-charge (m/z) ratios is according to ionizing molecules and then sorting and identifying the ions (Figure 4).

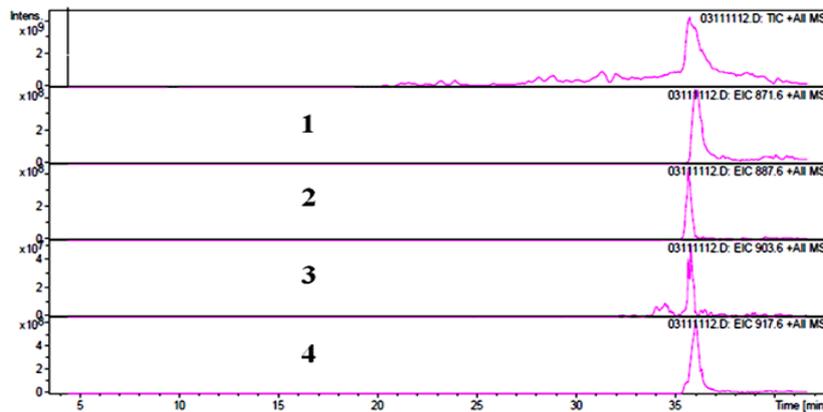


**Table 1:** LC-MS/MS Instrument Parameter and Feature Details.

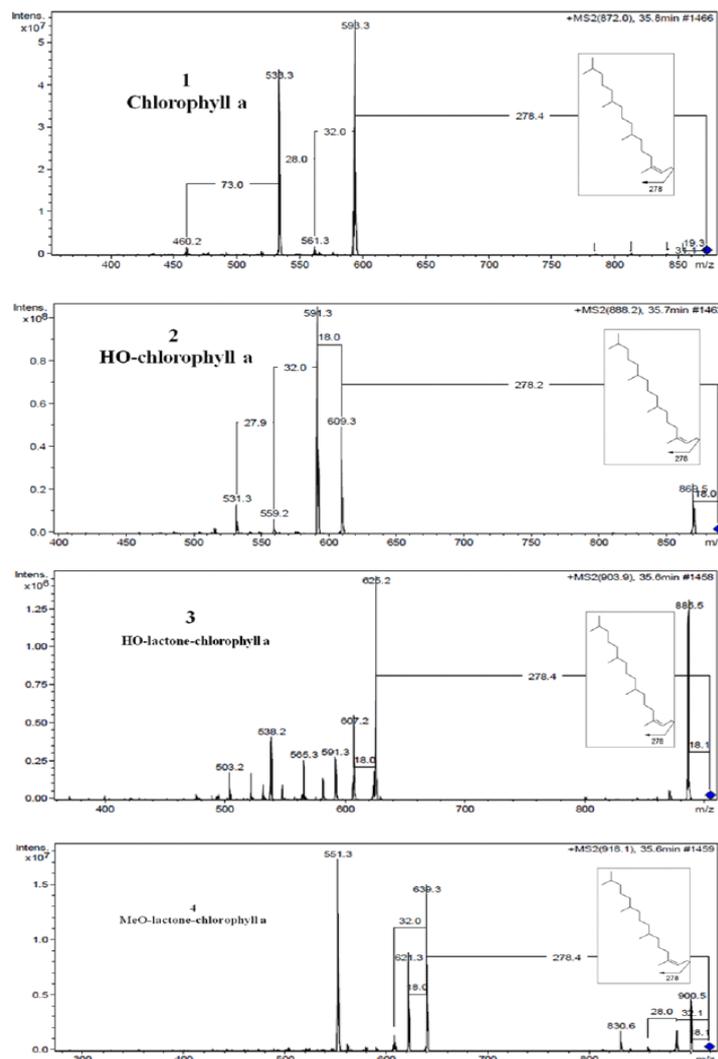
Parameter	Value
Dry Temp (°C)	350
Dry Gas (l/min)	8
Nebulizer (psi)	30
Capillary (V)	-5000
Skimmer (V)	85
Cap Exit (V)	300
Oct DC (V)	11.5
Oct 2 DC (V)	4.1
Trap Drive	144
Oct RF (Vpp)	300
Lens 1 (V)	-6.4
Lens 2 (V)	-76.4
Scan Range (m/z)	200-2200
Polarity (pos/neg)	Pos
Auto MS2	on
M/MS frag Ampl (V)	0.7

**Table 2:** molecular weight of protonated molecular ion' (MH+) of different chlorophyll a allomers before and after of separation of magnesium ion.

	Molecular weight in natural condition	Molecular weight of Mg	Molecular weight without of magnesium ion
Chlorophyll a	893	22	871
HO-chlorophyll a	909	22	887
HO-lactone-chlorophyll a	925	22	903
MeO-lactone-chlorophyll a	939	22	917



**Figure 3:** Total ion chromatogram (TIC) and Extracted ion chromatogram (EIC) of different chlorophyll  $\alpha$  allomers (1, Chlorophyll  $\alpha$  871.6  $m/z$ ); (2, HO-chlorophyll  $\alpha$  887.6  $m/z$ ); (3, HO-lactone-chlorophyll  $\alpha$  903.6  $m/z$ ) and (4, MeO-lactone-chlorophyll  $\alpha$  917.6  $m/z$ ) of the ASN\_M strain. The x-axis represents retention time (min), and the y-axis represents signal intensity. Intensity is measured in counts per second (cps).



**Figure 4:** MS/MS<sup>2</sup> fragmentation pattern of protonated chlorophyll  $\alpha$  allomers (chlorophyll  $\alpha$ ; 872  $m/z$ ), (HO-chlorophyll  $\alpha$ ; 888.2  $m/z$ ); (HO-lactone-chlorophyll  $\alpha$ ; 903.9  $m/z$ ) and (MeO-lactone-chlorophyll  $\alpha$ ; 918  $m/z$ ) of the ASN\_M strain.  $m/z$  = Mass-to-charge ratios, The intensity of the ion on the y-axis is given as counted ions per second (cps) and the mass-to-charge ratio ( $m/z$ ) on the x-axis

Liquid chromatography is a fundamental separation technique in the life sciences and related fields of chemistry. Unlike gas chromatography, which is unsuitable for nonvolatile and thermally fragile molecules, liquid chromatography can safely separate a very wide range of organic compounds, from small-molecule drug metabolites to peptides and proteins. Mass spectrometers work by ionizing molecules and then sorting and identifying the ions according to their mass-to-charge ( $m/z$ ) ratios. Two key components in this process are the ion source, which generates the ions, and the mass analyzer, which sorts the ions. Several different types of ion sources are commonly used for LC/MS. Each is suitable for different classes of compounds. Several different types of mass analyzers are also used [20].

## Conclusion

Here, we employed LC-MS to separate and identify the allomers of chlorophyll a produced in the *Nostoc* strain. Moreover, we extend the use of MS/MS<sup>2</sup> in the designation of allomers of chlorophyll a configuration to identification four different allomers during the methanolic allomerisation reactions of chlorophyll a. This research is the first documented of isolation four different chlorophyll a allomers of *Nostoc* sp. by liquid chromatography-mass spectrometer (LC-MS) in Iran.

## Acknowledgements

This study was designed and performed in Department of Biotechnology, Tehran, Iran and University of Helsinki, Department of Biology, Science and Research Branch. The authors would like to thank to Dr. David Fewer and Dr. Leo Rouhiainen of the Department of Food and Environment Sciences, University of Helsinki, for their helpful discussion, and also wish to thank, Lyudmila Saari for her/his helpful assistance.

## References

1. Airs RL, Temperton B, Sambles C, Farnham G, Skill SC and Llewellyn CA. Chlorophyll f and chlorophyll d are produced in the cyanobacterium *Chlorogloeopsis fritschii* when cultured under natural light and near-infrared radiation. *FEBS letters*. 2014 16;588(20):3770-3777.
2. Ritchie RJ. Universal chlorophyll equations for estimating chlorophylls a, b, c, and d and total chlorophylls in natural assemblages of photosynthetic organisms using acetone, methanol, or ethanol solvents. *Photosynthetica*. 2008;46(1):115-126.
3. Frei YF. The derivative absorption spectra of chlorophyll in algae and leaves at low temperatures. *Biochimica et biophysica acta*. 1962;57(1):82-87.
4. Boekema EJ, Hifney A, Yakushevskaya AE, Piotrowski M, Keestra W and Berry S, et al. A giant chlorophyll-protein complex induced by iron deficiency in cyanobacteria. *Nature*. 2001;412(6848):745-748.
5. Louda JW, Li J, Liu L, Winfree MN, Baker EW. Chlorophyll-degradation during cellular senescence and death. *Organic geochemistry*. 1998;29(5-7):1233-1251.
6. Liu L, Jokela J, Wahlsten M, Nowruzi B, Permi P and Zhang YZ, et al. Nostosins, trypsin inhibitors isolated from the terrestrial cyanobacterium *Nostoc* sp. strain FSN. *Journal of natural products*. 2014;77(8):1784-1790.
7. Nowruzi B, Ahmadimoghadam A. Two new records of heterocystus cyanobacteria (Nostocaceae) from paddy fields of Golestan Province. *Iran. Journ. Bot.* 2006;11(2):170-173.
8. Nowruzi B, Khavari-Nejad RA, Sivonen K, Kazemi B, Najafi F and Nejadstari T. Phylogenetic and morphological evaluation of two species of *Nostoc* (Nostocales, Cyanobacteria) in certain physiological conditions. *African Journal of Agricultural Research*. 2012 17;7(27):3887-3897.
9. Nowruzi B, Khavari-Nejad RA, Sivonen K, Kazemi B, Najafi F, Nejadstari T. A gene expression study on strains of *Nostoc* (Cyanobacteria) revealing antimicrobial activity under mixotrophic conditions. *African Journal of Biotechnology*. 2012;11(51):11296-11308.
10. Nowruzi B, Khavari-Nejad RA, Sivonen K, Kazemi B, Najafi F and Nejadstari T. Identification and toxigenic potential of a *Nostoc* sp. *Algae*. 2012;27(4):303-313.
11. Nowruzi B, Khavari-Nejad RA, Sivonen K, Kazemi B, Najafi F and Nejadstari T. Optimization of cultivation conditions to maximize extracellular investments of two *Nostoc* strains. *Algological Studies*. 2013;142(1):63-76.
12. Nowruzi B, Khavari-Nejad RA, Sivonen K, Kazemi B, Najafi F, Nejadstari T. Identification and toxigenic potential of a cyanobacterial strain (*Stigomena* sp.). *Progress in Biological Sciences*. 2013;3(1):79-85.
13. Nowruzi B, Khavari-Nejad RA, Nejadstari T, Sivonen K, Fewer D. A proposal for the unification of two cyanobacterial strains of *Nostoc* as the same species. *Rostaniha*. 2017;17(2):161-172.
14. Nowruzi B, Fahimi H, Ordodari N, Assareh R. Genetic analysis of polyketide synthase and peptide synthase genes of cyanobacteria as a mining tool for new pharmaceutical compounds. *Journal of Pharmaceutical & Health Sciences*. 2017;5(2):139-150.
15. Nowruzi B, Fahimi H, Ordodari N. Molecular phylogenetic and morphometric evaluation of *Calothrix* sp. N42 and *Scytonema* sp. N11. *Agriculture research education and extension organization*. 2017;18(2):210-221.
16. Nowruzi B, Haghhighat S, Fahimi H, Mohammadi E. *Nostoc* cyanobacteria species: a new and rich source of novel bioactive compounds with pharmaceutical potential. *Journal of Pharmaceutical Health Services Research*. 2018;9(1):5-12.
17. Nowruzi B, Blanco S. In silico identification and evolutionary analysis of candidate genes involved in the biosynthesis methylproline genes in cyanobacteria strains of Iran. *Phytochemistry Letters*. 2019;29:199-211.
18. Nowruzi B, Nejadstari T, Jokela J. Characterization of a New Peptide-Aldehyde Compound from the Terrestrial Cyanobacterium *Nostoc* sp. Bahar\_M by LC-MS and Marfey's analysis. *Iranian Journal of Biotechnology*. 2019;17(2):1563.
19. Walker JS, Jie C, Keely BJ. Identification of diastereomeric chlorophyll allomers by atmospheric pressure chemical ionisation liquid chromatography/tandem mass spectrometry. *Rapid communications in mass spectrometry*. 2003;17(11):1125-1131.
20. Heftmann E, editor. *Chromatography: Fundamentals and applications of chromatography and related differential migration methods-Part B: Applications*. Elsevier; 2004.